

COMBRETUM HEREROENSE SCHINZ (COMBRETACEAE): MEDICINAL USES, PHYTOCHEMISTRY, AND PHARMACOLOGICAL PROPERTIESALFRED MAROYI*

Department of Botany, University of Fort Hare, Alice, South Africa.

*Corresponding author: Alfred Maroyi; E-mail: amaroyi@ufh.ac.za

Received: 03 April 2025, Revised and Accepted: 25 July 2025

ABSTRACT

Traditional knowledge about *Combretum hereroense* Schinz has been poorly documented, despite the ethnobotanical studies conducted in various parts of tropical Africa. *C. hereroense* is a small deciduous to semi-deciduous tree widely used in traditional medicine throughout its distributional range in tropical Africa. The present review compiles existing information on the medicinal, phytochemical, and pharmacological properties of *C. hereroense*. A search for available information on the medicinal uses, phytochemical, and pharmacological properties of *C. hereroense* was conducted by systematically searching the scientific databases such as ScienceDirect®, PubMed®, Web of Science, SpringerLink®, Google Scholar, Scopus®, and SciELO, and as well as pre-electronic literature sources such as book chapters, books, and other scientific publications obtained from the university library. This study showed that the bark, fruit, leaf, root, stem, or shoot decoctions or infusions of *C. hereroense* are used as aphrodisiac, and traditional medicine against gastrointestinal problems, infertility in women, respiratory infections, sexually transmitted infections, bilharzia, general body pains, headache, heart diseases, heartburn, malaria, sores, and wounds. The phytochemical evaluation of the plant species revealed that it contains flavonoids, tannins, phenols, stilbenoids, phenanthrenes, and triterpenoids. The pharmacological evaluations showed that the crude extracts and phytochemical compounds isolated from the species have anthelmintic, antibacterial, antimycobacterial, antifungal, anti-inflammatory, and antioxidant activities. To realize the full potential of *C. hereroense* as a traditional medicine, future studies should focus on conducting detailed phytochemical, pharmacological, and toxicological evaluations, *in vivo* and clinical research.

Keywords: Bush willow, Combretaceae, *Combretum hereroense*, Traditional medicine, Tropical Africa.

© 2025 The Authors. Published by Innovare Academic Sciences Pvt Ltd. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>) DOI: <http://dx.doi.org/10.22159/ajpcr.2025v18i10.54438>. Journal homepage: <https://innovareacademics.in/journals/index.php/ajpcr>

INTRODUCTION

Combretum hereroense Schinz (Fig. 1) is a member of the Combretaceae family commonly known as the white mangrove, Indian almond, or bush willow family. *C. hereroense* is a multipurpose species, and these taxa are often associated with use categories such as food, medicines, fuel, timber, ornamental, recreational, and symbolic applications [1-5]. Unlike other *Combretum* species, *C. hereroense* serves as a source of security for communities in rural and peri-urban areas in providing firewood, traditional medicines, windbreaks, shade, and construction materials. Therefore, *C. hereroense* provides numerous ecosystem services and goods that are essential and support human well-being and survival [6-12]. Ecosystem services are the direct and indirect contributions of ecosystems to human well-being through supporting, provisioning, regulating, and cultural services [13-19]. *C. hereroense* is widely used in traditional medicine throughout its distributional range in tropical Africa [20,21]. *C. hereroense* is an important fodder species in tropical Africa, with its leaves, flowers, fruits, shoots, and young twigs being palatable and browsed by livestock and game, particularly during the dry periods [22-26]. *C. hereroense* is an attractive tree which is popular as an ornamental and/or shade plant in larger private gardens and also used as a street tree in tropical Africa, tolerating frost and moderate drought [22,26]. The wood of *C. hereroense* is brown in color, hard, heavy, strong, compact, easily worked, termite- and borer-proof, and makes a useful general-purpose timber as construction material for houses, livestock enclosures, fencing posts, handles of agricultural implements, furniture, and ornaments [22,26,27]. The wood of *C. hereroense* is also considered to be one of the most important and best quality firewoods as it burns slowly with intense heat, little smoke, and makes good charcoal [22,26,28,29]. The fruits and leaves of *C. hereroense* are used as a tea substitute in Botswana and Zimbabwe [22,23,25,26,30,31], but the seeds of the species are also reputed to be poisonous [25]. Therefore, this article reviews the

importance of *C. hereroense* in traditional medicine with a holistic approach that includes its botany, phytochemical, and pharmacological properties.

MATERIALS AND METHODS

A search for available information on medicinal uses, phytochemical, and pharmacological properties of *C. hereroense* was conducted by systematically searching the scientific databases such as ScienceDirect®, PubMed®, Web of Science, SpringerLink®, Google Scholar, Scopus®, and SciELO, and as well as pre-electronic literature sources such as book chapters, books, and other scientific publications obtained from the university library. The search was conducted from June 2024 to February 2025 using the following keywords: "*Combretum hereroense*," "biological activities of *Combretum hereroense*," "pharmacological properties of *Combretum hereroense*," "ethnobotany of *Combretum hereroense*," "medicinal uses of *Combretum hereroense*," "phytochemistry of *Combretum hereroense*," and "traditional uses of *Combretum hereroense*." The search covered publications from 1962 to 2024, a long period to capture literature on the medicinal, phytochemical, and pharmacological properties of *C. hereroense*.

RESULTS AND DISCUSSION**Botanical description of *C. hereroense***

The genus name "*Combretum*" is of classical origin, as the name was first used by the Roman naturalist, natural philosopher, naval and army commander Gaius Plinius Secundus, known in English as Pliny (23–79 AD), who used it in reference to an unknown plant [22,26]. The name was also reused by the Swedish botanist Pehr Löfving (31 January 1729–22 February 1756) for the *Combretum* genus [22,26]. The species name "*hereroense*" means "from Hereroland" or "inhabited by the Herero people" [32], and this name is in honor of the Herero people of Namibia [22]. The common name 'bush willow'

indicates a superficial resemblance of the species to willows, that is, species belonging to the genus *Salix* L. (family Salicaceae), but “bush willows” and “willows” are not closely related to each other [22]. Other common names of the species include “mouse-eared bush willow,” “mouse-eared combretum,” and “russet bush willow” [22,24-26]. The synonyms of *C. hereroense* Schinz include *C. borumense* Engl. and Diels, *C. bruchhausenianum* Engl. and Diels, *C. cufodontii* Chiov., *C. eilkerianum* Schinz, *C. erlangerianum* Engl. and Diels, *C. greenwayi* Exell, *C. grotei* Exell, *C. hereroense* Schinz var. *villosissimum* Engl. and Diels, *C. parvifolium* Engl., *C. porphyrolepis* Engl. and Diels, *C. rautanenii* Engl. and Diels, *C. rhodesiacum* Baker f., *C. sambesiaceum* Engl. and Diels, *C. transvaalesnse* Schinz, *C. transvaalesnse* Schinz var. *ochrolepidotum* Dümmer, *C. transvaalesnse* Schinz var. *villosissimum* (Engl. and Diels) Burt Davy, *C. usaramense* Engl., *C. villosissimum* (Engl. and Diels) Engl. and *C. volkensii* Engl. [22,24,33-41]. Four infraspecific taxa of *C. hereroense* (Table 1) are recognized, and these are *C. hereroense* Schinz subsp. *grotei* (Exell) Wickens, *C. hereroense* Schinz subsp. *hereroense*, *C. hereroense* Schinz var. *parvifolium* (Engl.) Wickens and *C. hereroense* Schinz subsp. *volkensii* (Engl.) Wickens [33-44]. Ethnopharmacological and ethnobotany research rarely makes any reference to these infraspecific taxa; therefore, *C. hereroense sensu lato* is adopted throughout the manuscript.

C. hereroense is a small semi-deciduous to deciduous tree growing up to 11 m in height [22,24,26]. The species is usually single or many-stemmed (Fig. 1a), the trunk often crooked, up to about 23 cm in diameter, with a light or dark brown, thick, rough, fissured bark, which is dark grey in color. The bark on young branches and branchlets is stringy, hairy, and peeling in strips, but rough and longitudinally fissured on older branches and stems [22,26]. The leaves are opposite to sub-opposite, simple, coarse-textured, on short lateral twigs, narrowly to broadly elliptic or broadly obovate, widest about or above

the middle, varying from glabrous to grey, long-hairy above and, below, from densely golden brown scaly to green with only a few scales on the veins. The leaves often turn red-brown late in the rainy season, apex and base are broadly tapering to rounded, with tertiary veins that are notably raised below. The flowers are greenish-yellow (Fig. 1b), sweet-scented, borne in dense axillary spikes, and appear before the new leaves. The fruits are winged (Fig. 1c), brown in color, borne in such abundance that they weigh down the branches, and strikingly rich, deep brown between the wings. *C. hereroense* has been recorded in Angola, Botswana, Eswatini, Ethiopia, Kenya, Malawi, Mozambique, Namibia, Somalia, South Africa, South Sudan, Sudan, Tanzania, Uganda, Zambia, and Zimbabwe [22,33-49] (Fig. 2). *C. hereroense* has been recorded in low to medium altitudes in woodland, bushveld, grassland, wooded grassland, in stony, shallow or brack soil, in sandy or red loam, on hillsides or valleys, sometimes on stream banks, termite mounds and on Vlei margins, often several together at an altitude ranging from 30 m to 1465 m above sea level [22,24,37,40].

Ethnomedicinal applications of *C. hereroense*

C. hereroense is used as a source of traditional medicines in South Africa, Eswatini, Angola, Botswana, Kenya, Tanzania, Somalia, Namibia, and Zimbabwe, that is, 56.3% of the countries where the species is indigenous (Table 2). In South Africa, the roots of *C. hereroense* are sold in informal herbal medicine markets as sources of traditional medicines [50]. *C. hereroense* is also categorized as a valuable medicinal plant species in South Africa, as the species is used by the majority of cultural groups in that country. The species has been incorporated into the traditional *materia medica* of that country, and *C. hereroense* is included in two South African encyclopedias, namely, “medicinal and magical plants of southern Africa: An annotated checklist” [51] and “medicinal plants of South Africa” [52]. These two monographs documented the botanical descriptions of medicinal plants that are regarded as major components of the South African *materia medica*, the plant parts used, medicinal applications, preparation, dosage, active ingredients, and pharmacological properties of the species [51,52].

The traditional medicines prepared from the bark, fruit, leaf, root, stem, or shoot decoctions or infusions of *C. hereroense* are used to treat and manage 34 human and animal diseases and ailments (Table 1). The main ailments and diseases treated by *C. hereroense* extracts (Fig. 3) include the use of roots as aphrodisiac, and use of the bark, fruit, leaf, root, stem, or shoot decoctions or infusions as traditional medicine against gastro-intestinal problems, infertility in women, respiratory infections, sexually transmitted infections, bilharzia, general body pains, headache, heart diseases, heartburn, malaria, sores and wounds. In Kenya, the roots of *C. hereroense* are mixed with those of *Uvaria leptoclados* Oliv. (Annonaceae family) as a remedy for erectile dysfunction [53,54], menstrual problems [53], postpartum [53], and retained placenta [53]. Similarly, the roots of *C. hereroense* are mixed with those of *Balanites aegyptiaca* (L.) Delile (Zygophyllaceae family) is a traditional medicine for infertility in women [55].

Phytochemistry and pharmacological properties of *C. hereroense*

Qualitative and quantitative phytochemical analyses of *C. hereroense* fruits and leaves revealed the presence of flavonoids, tannins,

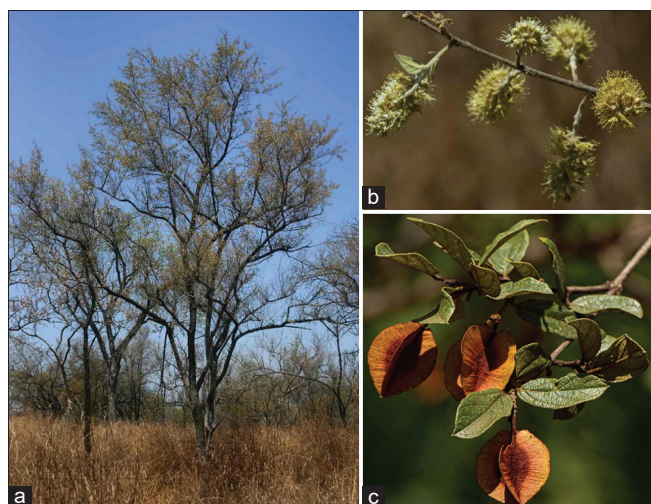


Fig. 1: *Combretum hereroense*: (a) Entire plant, (b) branch showing flowers, and (c) branch showing leaves and fruits (photos: BT Wursten)

Table 1: Main morphological characters of *Combretum hereroense* subspecies

Subspecies	Key morphological features	Distribution	References
<i>grotei</i>	Leaves <2.0 cm long, fruits 1.5–2.0 cm long and scales on leaf contiguous	Kenya, South Sudan, Sudan, Tanzania and Uganda	[33,34,42,43]
<i>hereroense</i>	Leaves >2.0 cm long, fruits >2.0 cm long, and scales on the leaf contiguous	Angola, Botswana, Namibia, Kenya, South Africa, Malawi, Mozambique, Eswatini, Tanzania, Zambia, Zimbabwe	[33,34,40,44]
<i>parvifolium</i>	Leaves pubescent, fruits 2.5 cm long, and scales on the leaf not impressed	Ethiopia, Kenya, and Tanzania	[33,34,36]
<i>volkensii</i>	Leaves glabrous, up to 8.0 cm long, fruits 1.5–2.5 cm long, scales on leaf impressed and not contiguous	Kenya, Somalia, Tanzania, and Uganda	[33,34,42]



Fig. 2: Distribution of *Combretum hereroense* in tropical Africa

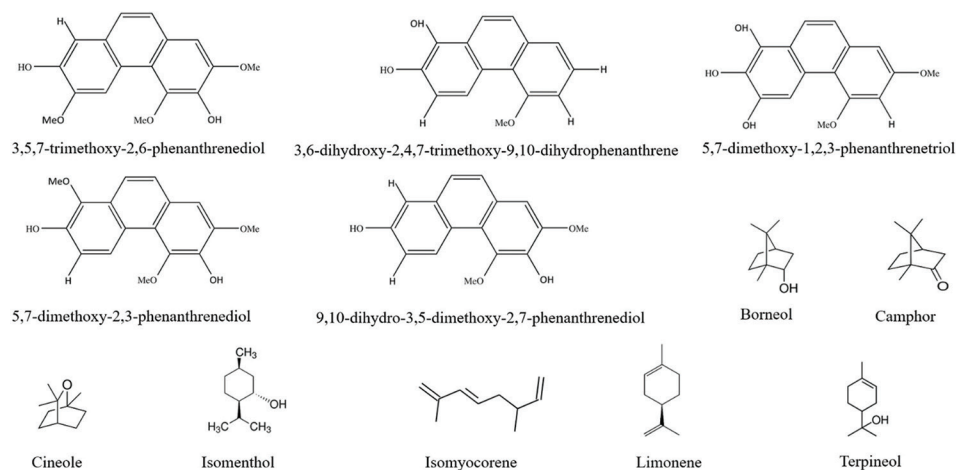


Fig. 3: Chemical structures of phytochemical compounds isolated from *Combretum hereroense*

phenols, stilbenoids, phenanthrenes, and triterpenoids [88-90] (Table 3 and Fig. 3). Some of the phytochemical compounds isolated from *C. hereroense* and its crude extracts exhibited anthelmintic, antibacterial, antimycobacterial, antifungal, anti-inflammatory, and antioxidant activities.

Anthelmintic activities

McGaw *et al.* [91] evaluated the anthelmintic activities of acetone extracts of *C. hereroense* leaves against the free-living nematode

Caenorhabditis elegans var. Bristol (N2) with the standard nematocidal drug levamisole as a positive control. The extract exhibited activities against the nematode [91].

Anti-bacterial activities

Cock and Van Vuuren [68] evaluated the antibacterial activities of aqueous and methanol extracts of *C. hereroense* leaves against *Alicigenes faecalis*, *Aeromonas hydrophilia*, *Bacillus cereus*, *Bacillus subtilis*, *Citrobacter freundii*, *Escherichia coli*, *Klebsiella pneumonia*,

Table 2: Medicinal uses of *Combretum hereroense*

Medicinal use	Part used	Country	References
Mono-therapeutic applications			
Aphrodisiac	Root decoction taken orally	Botswana and Zimbabwe	[30,56]
Bilharzia	Leaf, root, or shoot decoction taken orally	Tanzania	[21,57-64]
Gastrointestinal problems (diarrhea, dysentery, and stomach problems)	Leaf, root, shoot, or young stem decoction or infusion taken orally	Eswatini, Namibia, South Africa, Tanzania, and Zimbabwe	[21,22,26,31,32,52, 56,57,61,62-70]
General body pains	Leaf or root decoction or infusion taken orally	Angola, Botswana, and South Africa	[32,59,66,71]
Headache	Fruit, leaf, root, or shoot decoction taken orally	Botswana, South Africa, Tanzania, and Zimbabwe	[21,57,58,60,61,63,64,66,72]
Heart diseases	Bark decoction taken orally	South Africa	[21,31,32,66,73,74]
Heartburn	Bark decoction taken orally	South Africa	[21,31,32,66,73,74]
Infertility in women	Bark or root powder taken orally	Botswana, South Africa, and Zimbabwe	[52,56,58,60,63,64,66,68,70,72]
Leprosy	Not specified	Not specified	[64]
Malaria	Not specified	South Africa	[64,69]
Penal swellings	Root decoctions applied topically	Botswana	[59]
Respiratory infections (bronchial problems, chest pains, cold, cough, loss of voice, sore throat, tonsillitis, and tuberculosis)	Bark, fruit, leaf, root, shoot, or stem decoction or infusion taken orally	Botswana, Kenya, Namibia, Somalia, South Africa, Tanzania, Zambia, and Zimbabwe	[21,30,31,52, 57,61,63,66-68,70,75-85]
Sexually transmitted infections (chlamydia, gonorrhea, and venereal disease)	Leaf, root, or young stem decoction or infusion taken orally	Botswana, Namibia, South Africa, Zambia, and Zimbabwe	[21,32,52,59,64,67,68,70, 75,85,86]
Skin softening cream	Root decoction taken orally	Botswana	[30]
Sores and wounds	Not specified	South Africa	[52,64,68,70]
Tonic	Not specified	South Africa	[69]
Ulcers	Not specified	Not specified	[64]
Vulval swellings	Root decoctions applied topically	Botswana	[59]
Ethnoveterinary medicine (constipation, cough, dysentery, and pain)	Leaves	South Africa	[87]
Used in combination with other species			
Erectile dysfunction	Roots mixed with those of <i>Uvaria leptacladon</i> Oliv. (Annonaceae family)	Kenya	[53,54]
Infertility in women	Roots mixed with those of <i>Balanites aegyptiaca</i> (L.) Delile (Zygophyllaceae family)	Kenya	[55]
Menstrual problems, postpartum, and retained placenta	Roots mixed with those of <i>U. leptacladon</i> Oliv. (Annonaceae family)	Kenya	[53]

Table 3: Phytochemical composition of *Combretum hereroense*

Phytochemical compound	Formula	Part	References
3,5,7-trimethoxy-2,6-phenanthrenediol	C ₁₇ H ₁₆ O ₅	Fruits	[89]
3,6-dihydroxy-2,4,7-trimethoxy-9,10-dihydrophenanthrene	C ₁₇ H ₁₈ O ₄	Leaves	[88]
5,7-dimethoxy-1,2,3-phenanthrenetriol	C ₁₆ H ₁₄ O ₅	Fruits	[89]
5,7-dimethoxy-2,3-phenanthrenediol	C ₁₆ H ₁₄ O ₄	Fruits	[89]
9,10-dihydro-3,5-dimethoxy-2,7-phenanthrenediol	C ₁₆ H ₁₆ O ₄	Fruits	[89]
Borneol	C ₁₀ H ₁₈ O	Leaves	[90]
Camphor	C ₁₀ H ₁₆ O	Leaves	[90]
Cineole	C ₁₀ H ₁₈ O	Leaves	[90]
Isomenthol	C ₁₀ H ₂₀ O	Leaves	[90]
Isomyocorene	C ₁₀ H ₁₆	Leaves	[90]
Limonene	C ₁₀ H ₁₆	Leaves	[90]
Terpineol	C ₁₀ H ₁₈ O	Leaves	[90]

Proteus mirabilis, *Proteus vulgaris*, *Pseudomonas aeruginosa*, *Pseudomonas fluorescens*, *Salmomnellatyphimurin*, *Serratiamarcescens*, *Shigella sonnei*, *Stapylococcus aureus*, and *Staphylococcus epidermidis* using the disc diffusion assay. The extracts exhibited activities against the tested pathogens with minimum inhibition concentration (MIC) values ranging from 337.0 µg/ml to 1440.0 µg/ml [68]. Katerere *et al.* [89] evaluated the antibacterial activities of the phytochemical compounds 5,7-dimethoxy-1,2,3-phenanthrenetriol, 5,7-dimethoxy-2,3-phenanthrenediol, 9,10-dihydro-3,5-dimethoxy-2,7-

phenanthrenediol, and 3,5,7-trimethoxy-2,6-phenanthrenediol isolated from *C. hereroense* fruits against *Mycobacterium fortuitum*, *Proteus vulgaris*, and *Staphylococcus aureus* using the microtiter dilution assay with streptomycin as a positive control. The phytochemical compounds exhibited activities against the tested pathogens with MIC values ranging from 25.0 µg/ml to 100.0 µg/ml [89]. Grimsey *et al.* [90] evaluated the antibacterial activities of methanol extracts of *C. hereroense* against *Staphylococcus aureus*, *Klebsiella pneumoniae*, and *Escherichia coli* using the broth

microdilution assay with ciprofloxacin, ampicillin, gentamycin, oxacillin, and methicillin as positive controls. The extracts exhibited activities against the tested pathogens with MIC values ranging from 813.0 µg/mL to 1625.0 µg/mL [90]. Eloff [92] evaluated the antibacterial activities of acetone extracts of *C. hereroense* leaves against *Escherichia coli*, *Staphylococcus aureus*, *Enterococcus faecalis*, and *Pseudomonas aeruginosa* using the twofold serial dilution with gentamycin as a positive control. The extracts exhibited activities against the tested pathogens with MIC values ranging from 0.2 mg/mL to 6.0 mg/mL [92]. Fyhrquist *et al.* [93] evaluated the antibacterial activities of methanol extracts of *C. hereroense* leaves against *Sarcina* spp. and *Staphylococcus epidermidis* using the agar diffusion method with ampicillin and streptomycin as positive controls. The extract exhibited activities against *Sarcina* spp. and *Staphylococcus epidermidis* with MIC values of 23.3 mg/ml and 30.0 mg/ml, respectively [93]. Anokwuru *et al.* [94] evaluated the antibacterial activities of methanol extracts of *C. hereroense* leaves against *Staphylococcus aureus*, *Staphylococcus epidermidis*, *Bacillus cereus*, *Klebsiella pneumoniae*, *Enterococcus faecalis*, *Pseudomonas aeruginosa*, *Escherichia coli*, *Salmonella typhimurium*, and *Shigella sonnei* using the microdilution assay with ciprofloxacin as a positive control. The extracts exhibited activities against the tested pathogens with MIC values ranging from 0.37 mg/mL to >3.0 mg/mL [94].

Antimycobacterial activities

Masoko and Nxumalo [95] evaluated the antimycobacterial activities of dichloromethane, hexane, acetone, and methanol extracts of *C. hereroense* leaves against *Mycobacterium smegmatis* using the microdilution assay with rifampicin as a positive control. The extracts exhibited activities against the tested pathogen with MIC values ranging from 0.5 mg/ml to 1.9 mg/ml [95]. Komape *et al.* [96] evaluated the antimycobacterial activities of dichloromethane, hexane, acetone and methanol extracts of *C. hereroense* leaves against *Mycobacterium smegmatis* using the serial microplate dilution method with rifampicin as a positive control. The extracts exhibited activities against the tested pathogen with MIC values ranging from 0.6 mg/ml to 1.6 mg/ml [96].

Antifungal activities

Cock and Van Vuuren [68] evaluated the antifungal activities of aqueous and methanol extracts of *C. hereroense* leaves against *Aspergillus niger*, *Candida albicans*, and *Rhizopus stolonifer* using the disc diffusion assay. The extracts exhibited activities against the tested pathogens with MIC values ranging from 287.0 µg/ml to 5567.0 µg/ml [68]. Katerere *et al.* [89] evaluated the antifungal activities of the phytochemical compounds 5,7-dimethoxy-1,2,3-phenanthrenetriol, 5,7-dimethoxy-2,3-phenanthrenediol, 9,10-dihydro-3,5-dimethoxy-2,7-phenanthrenediol, and 3,5,7-trimethoxy-2,6-phenanthrenediol isolated from *C. hereroense* fruits against *Candida albicans* using the microtiter dilution assay with fluconazole as a positive control. The phytochemical compounds exhibited activities against the tested pathogen with an MIC value of 50.0 µg/ml [89]. Masoko *et al.* [97] evaluated the antifungal activities of hexane, acetone, methanol, and dichloromethane extracts of *C. hereroense* leaves against *Candida albicans*, *Aspergillus fumigatus*, *Cryptococcus neoformans*, *Sporothrix schenckii*, and *Microsporum canis* using the microdilution assay with amphotericin B as a positive control. The extracts exhibited activities against the tested pathogens with MIC values ranging from 0.02 mg/ml to 2.5 mg/ml [97].

91 Anti-inflammatory activities

McGaw *et al.* [91] evaluated the anti-inflammatory activities of water, acetone, and ethyl acetate extracts of *C. hereroense* leaves in an *in vitro* assay for cyclooxygenase inhibitors with indomethacin as a positive control. The extracts exhibited activities by showing cyclooxygenase inhibition ranging from 70.0% to 81.0% [91]. Eloff *et al.* [98] evaluated the anti-inflammatory activities of the acetone extract of *C. hereroense* leaves using the radiochemical cyclooxygenase bioassay against sheep seminal vesicles. The extract showed from 76.0% to 84.0% inhibition of cyclooxygenase [98].

Antioxidant activities

Masoko and Eloff [99] evaluated the antioxidant activities of acetone and methanol extracts of *C. hereroense* leaves using the 2,2-diphenyl-1-picryl hydrazyl-free radical scavenging assay. The extract exhibited moderate antioxidant activities [99].

CONCLUSION

The current review provides a summary of the medicinal uses, phytochemical, and pharmacological properties of *C. hereroense*. However, there is a need for detailed studies focusing on phytochemical and pharmacological properties, toxicity and safety, mechanisms of action *in vivo*, and clinical research of the species aimed at corroborating the ethnomedicinal applications of *C. hereroense*. There is also a need for ethnopharmacological studies aimed at examining the combinational effects of *C. hereroense* extracts with other plant species such as *B. aegyptiaca* and *U. leptacladon*.

AUTHOR CONTRIBUTION

AM conceptualized the research and wrote the manuscript.

CONFLICT OF INTEREST

No conflict of interest is associated with this research.

AUTHOR FUNDING

None.

REFERENCES

1. Etefa G, Antony JR, Kidane G, Alemayehu T. Domestication of indigenous fruit and fodder trees/shrubs in dryland agroforestry and its implication on food security. *Int J Ecosys*. 2014;4:83-8. doi: 10.5923/j.ije.20140402.06.
2. Mensah S, Veldtman R, Assogbadjo AE, Ham C, Glèlè Kakaï RG, Seifert T. Ecosystem service importance and use vary with socio-environmental factors: A study from household-surveys in local communities of South Africa. *Ecosyst Serv*. 2017;23:1-8. doi: 10.1016/j.ecoser.2016.10.018.
3. Mao S, Shen Y, Deng H. Multipurpose plant utilization in ethnic areas of Guizhou, southwest China. *Ecol Indic*. 2018;90:547-53. doi: 10.1016/j.ecolind.2018.03.064.
4. Lelamo LL. A review on the indigenous multipurpose agroforestry tree species in Ethiopia: management, their productive and service roles and constraints. *Heliyon*. 2021;7(9):e07874. doi: 10.1016/j.heliyon.2021.e07874, PMID 34504969.
5. Allegrini A, Salvaneschi P, Schirone B, Cianfaglione K, Di Michele A. Multipurpose plant species and circular economy: *Corylus avellana* L. as a study case. *Front Biosci (Landmark Ed)*. 2022;27(1):11. doi: 10.31083/j.fb12701011, PMID 35090316.
6. Ouédraogo I, Nacoulma BM, Hahn K, Thiombiano A. Assessing ecosystem services based on indigenous knowledge in southeastern Burkina Faso (West Africa). *Int J Biodivers Sci Ecosyst Serv Manag*. 2014;10(4):313-21. doi: 10.1080/21513732.2014.950980.
7. Bidak LM, Kamal SA, Halmi MW, Heneidy SZ. Goods and services provided by native plants in desert ecosystems: examples from the northwestern coastal desert of Egypt. *Glob Ecol Cons*. 2015;3:433-47. doi: 10.1016/j.gecco.2015.02.001.
8. Robi MK, Edris EM. Distribution, abundance, and population status of four indigenous threatened tree species in the Arba Minch natural forest, southern Ethiopia. *Int J Nat Res Ecol Manag*. 2017;2:1-8. doi: 10.11648/j.ijnrem.20170201.11.
9. Ahammad R, Stacey N, Sunderland TC. Use and perceived importance of forest ecosystem services in rural livelihoods of Chittagong Hill Tracts, Bangladesh. *Ecosyst Serv*. 2019;35:87-98. doi: 10.1016/j.ecoser.2018.11.009.
10. Thorn JP, Thornton TF, Helfgott A, Willis KJ. Indigenous uses of wild and tended plant biodiversity maintain ecosystem services in agricultural landscapes of the Terai Plains of Nepal. *J Ethnobiol Ethnomed*. 2020;16(1):33. doi: 10.1186/s13002-020-00382-4, PMID 32513199.
11. Kimpouni V, Nzila JD, Watha-Ndoudy N, Madzella-Mbiemo MI, Yallo Mouhamed SY, Kampe JP. Exploring local people's perception of ecosystem services in Djoumouna Periurban Forest, Brazzaville, Congo. *Int J For Res*. 2021;2021:Article ID 6612649.

- doi: 10.1155/2021/6612649.
12. Maroyi A. Traditional uses of wild and tended plants in maintaining ecosystem services in agricultural landscapes of the Eastern Cape Province in South Africa. *J Ethnobiol Ethnomed*. 2022;18(1):17. doi: 10.1186/s13002-022-00512-0, PMID 35292046. Braat LC, de Groot R. The ecosystem services agenda: bridging the worlds of natural science and economics, conservation and development, and public and private policy. *Ecosyst Serv*. 2012;1(1):4-15. doi: 10.1016/j.ecoser.2012.07.011.
 14. Hasan SS, Zhen L, Miah MG, Ahamed T, Samie A. Impact of land use change on ecosystem services: a review. *Environ Dev*. 2020;34:100527. doi: 10.1016/j.envdev.2020.100527.
 15. Aristianti N, Aswad M, Arsyad A, Nursamsiar N, Nur S, Islam AA. The potential combination of *Centella asiatica*, *Curcuma longa*, and *Piper nigrum* extracts in treating brain injury: *in vitro*, *in vivo* and silico studies. *Int J App Pharm*. 2025;17(2):174-89. doi: 10.22159/ijap.2025v17i2.53173.
 16. Baba H, Bunu SJ. Spectroscopic and molecular docking analysis of phytoconstituent isolated from *Solenostemon monostachyus* as potential cyclooxygenase enzymes inhibitor. *Int J Chem Res*. 2025;9(1):1-6. doi: 10.22159/ijcr.2025v9i1.241.
 17. Bharti N, Yadav P, Kansotiya AK, Mali PC. Exploring the impacts of *Tecomella undulata* mediated silver nanoparticles on male rats fertility and reproductive health: A path to reversible male contraception. *Int J Curr Pharm Res*. 2025;17(1):83-9. doi: 10.22159/ijcpr.2025v17i1.6037.
 18. Yudianti Mendra NN, Suradnyana IG, Kusuma Wardani IG, Vernandes Sasadara MM. Nanoemulsion of champaca flower (*Magnolia alba*) oil as an antibacterial candidate: optimization, characterization, and thermodynamic stability testing. *Int J App Pharm*. 2025;17(2):115-25. doi: 10.22159/ijap.2025v17i2.53016.
 19. Rani R, Gahirwar A, Yadav S. Medicinal plants from the Himalayas: insights into their anticancer effects. *Int J Pharm Pharm Sci*. 2025;17(2):16-25. doi: 10.22159/ijpps.2025v17i2.52819.
 20. Neuwinger HD. African traditional medicine. Stuttgart: MedPharm Scientific Publishers; 2000.
 21. Quattrocchi U. CRC World dictionary of medicinal and poisonous plants: common names, scientific names, eponyms, synonyms, and etymology. Boca Raton: CRC Press; 2012.
 22. Palmer E, Pitman N. Trees of southern Africa, covering all known indigenous species in the Republic of South Africa, South-West Africa, Botswana, Lesotho and Swaziland. Cape Town: A A Balkema Publishers; 1972.
 23. McGregor J. Woodland resources: ecology, policy and ideology. An historical case study of woodland use in Shurugwi communal area, Zimbabwe. Loughborough [PhD thesis], Loughborough University of Technology; 1991.
 24. Palgrave MC. Keith Coates Palgrave trees of southern Africa. Cape Town: Struik Publishers; 2002.
 25. Van Wyk BE, Van Wyk P. Field guide to trees of southern Africa. Cape Town: Struik Nature; 2013.
 26. Venter F, Venter JA. Making the most of indigenous trees. Pretoria: Briza Publications; 2015.
 27. Shopo B, Mapaya RJ, Maroyi A. The traditional uses of plant diversity in Gokwe South District, Zimbabwe: timber and construction, ethnoveterinary medicine, firewood and charcoal, food, tools and handicraft, religious ceremonies and rituals and other uses. *Ethnobot Res Appl*. 2022;24:34. doi: 10.32859/era.24.34.1-23.
 28. Raseth MT, Semenya SS, Potgieter MJ, Maroyi A. The utilization and management of plant resource in rural areas of the Limpopo province, South Africa. *J Ethnobiol Ethnomed*. 2013;9:27. doi: 10.1186/1746-4269-9-27, PMID 23590903.
 29. Van Wyk BE, Gericke N. People's plants: A guide to useful plants of southern Africa. Pretoria: Briza Publications; 2018.
 30. Heath A, Heath R. Field guide to the plants of northern Botswana including the Okavango Delta. Royal Botanic Gardens, Richmond, Kew; 2009.
 31. Malatji RM. *Combretum hereroense* Schinz. 2019 Available. [cited Nov 13 2024] Available from: <https://pza.sanbi.org/combretum-hereroense>.
 32. Schmidt E, Lotter M, McClelland W. Trees and shrubs of Mpumalanga and Kruger National Park. Johannesburg: Jacana Media; 2017; 2007.
 33. Wickens GE. The *Combretum hereroense* complex (Combretaceae). *Kew Bull*. 1971;25(3):413-16. doi: 10.2307/4103188.
 33. Figueiredo E, Smith GF. Plants of Angola. Pretoria: Strelitzia. Vol. 22. National Botanical Institute; 2008.
 34. Wickens GE. Combretaceae: flora of tropical East Africa. London: Crown Publishing Group Agents for Oversea Governments and Administrations; 1973.
 35. Exell AW. Combretaceae. In: Launert E, editor. Flora Zambesiaca volume 4. Richmond: Kew Publishing and Flora Zambesiaca Managing Committee; 1978. p. 100-83.
 36. Edwards S, Tadesse M, Hedberg I. Flora of Ethiopia and Eritrea volume 2 part 2. Addis Ababa: the national herbarium. Addis Ababa University; 1995.
 37. Germishuizen G, Meyer NL. Plants of southern Africa: an annotated checklist. Pretoria: Strelitzia. Vol. 14. National Botanical Institute; 2003.
 39. Mannheimer CA, Curtis BA. Le Roux and Müller's field guide to the trees and shrubs of Namibia. Windhoek: Macmillan Education Namibia; 2009.
 40. Jordaan M, Van Wyk AE, Maurin O. A conspectus of *Combretum* (Combretaceae) in southern Africa, with taxonomic and nomenclatural notes on species and sections. *Bothalia*. 2011;41(1):135-60. doi: 10.4102/abc.v41i1.36.
 41. Burrows J, Burrows S, Lötter M, Schmidt E. Trees and shrubs of Mozambique. Cape Town: publishing print matters; 2018.
 42. Kalema J, Beentje H. Conservation checklist of the trees of Uganda. Richmond: Kew Publishing, Royal Botanic Gardens; 2012.
 43. Darbyshire I, Kordofani M, Farag I, Candiga R, Pickering H. The plants of Sudan and South Sudan. Richmond: Kew publishing, Royal Botanic Gardens; 2015.
 44. Govaerts R, Nic Lughadha E, Black N, Turner R, Paton A. The world checklist of vascular plants, a continuously updated resource for exploring global plant diversity. *Sci Data*. 2021;8(1):215. doi: 10.1038/s41597-021-00997-6, PMID 34389730.
 45. Drummond RB. A list of trees, shrubs and woody climbers indigenous or naturalised in Rhodesia. Kirkia. 1975;10:229-89.
 46. Setshogo MP, Venter F. Trees of Botswana: names and distribution. Pretoria: Southern African Botanical Diversity Network Report No. 18. SABONET. National Botanical Institute; 2005.
 47. Mapaura A, Timberlake J. A checklist of Zimbabwean vascular plants. Pretoria: Southern African Botanical Diversity Network Report No. 33. SABONET; 2004.
 48. Loffler L, Loffler P. Swaziland tree atlas: including selected shrubs and climbers. Pretoria: Southern African Botanical Diversity Network Report No. 38. SABONET; 2005; 2005.
 49. Setshogo MF. Preliminary checklist of the plants of Botswana. Pretoria: Southern African Botanical Diversity Network Report No. 37. SABONET. National Botanical Institute; 2005.
 50. Williams VL, Balkwill K, Witkowski ET. A lexicon of plants traded in the Witwatersrand umuthi shops, South Africa. *Bothalia*. 2001;31(1):71-98. doi: 10.4102/abc.v31i1.508.
 51. Arnold TH, Prentice CA, Hawker LC, Snyman EE, Tomalin M, Crouch NR, *et al*. Medicinal and magical plants of southern Africa: an annotated checklist. Pretoria: National Botanical Institute; 2002.
 52. Van Wyk BE, Van Oudtshoorn B, Gericke N. Medicinal plants of South Africa. Pretoria: Briza Publications; 2013.
 53. Kaingu CK, Oduma JA, Mbaria JM, Kiama SG. Medicinal plants traditionally used for the management of female reproductive health dysfunction in Tana River County, Kenya. *TANG [HUMANITAS MEDICINE]*. 2013;3(2):17.1-17.10. doi: 10.5667/tang.2013.0006.
 54. Kyarimpa C, Nagawa CB, Omara T, Odongo S, Ssebugere P, Lugasi SO *et al*. Medicinal plants used in the management of sexual dysfunction, infertility and improving virility in the East African Community: A systematic review. *Evid Based Complement Alternat Med*. 2023;2023:Article ID 6878852. doi: 10.1155/2023/6878852, PMID 37600549.
 55. Kiptisia RT, Nandwa AN. Phytochemical screening and acute oral toxicity study of root extracts of *Combretum hereroense* Schinz and *Balanites aegyptiaca* Del. traditionally used to treat female infertility in Baringo County, Kenya. *Eur J Med Pl*. 2023;34(1):12-22. doi: 10.9734/EJMP/2023/v34i11116.
 56. Shopo B, Mapaya RJ, Maroyi A. Ethnobotanical study of medicinal plants traditionally used in Gokwe South District, Zimbabwe. *S Afr J Bot*. 2022;149:29-48. doi: 10.1016/j.sajb.2022.05.052.
 57. Watt JM, Breyer-Brandwijk MG. The medicinal and poisonous plants of southern and eastern Africa being an account of their medicinal and other uses, chemical composition, pharmacological effects and toxicology in man and animal. Edinburgh: E and S. Livingstone Limited; 1962.
 58. Gelfand M, Mavi S, Drummond RB, Ndemera B. The traditional medical practitioner in Zimbabwe: his principles of practice and pharmacopoeia. Gweru: Mambo Press; 1985.
 59. Hedberg I, Staugard F. Traditional medicinal plants in Botswana. Gaborone: Ipeleng Publishers; 1989.
 60. Rogers CB, Verota L. Chemistry and biological properties of the African Combretaceae. In: Hostettmann K,

- Chinyanganya F, Maillard M, Wolfender JL, editors. Chemistry, biological and pharmacological properties of African medicinal plants. Harare: University of Zimbabwe Publications; 1996. p. 121-42.
61. Fyhrquist P, Mwasumbi L, Hægström CA, Vuorela H, Hiltunen R, Vuorela P. Antifungal activity of selected species of *Terminalia*, *Pteleopsis* and *Combretum* (Combretaceae) collected in Tanzania. *Pharm Biol.* 2004;42(4-5):308-17. doi: 10.1080/13880200490511891.
 62. Kokwaro JO. Medicinal plants of East Africa. Nairobi: University of Nairobi Press; 2009.
 63. Magwenzi R, Nyakunu C, Mukanganyama S. The effect of selected combretum species from Zimbabwe on the growth and drug efflux systems of *Mycobacterium aurum* and *Mycobacterium smegmatis*. *J Microb Biochem Technol.* 2014;3:3. doi: 10.4172/1948-5948.S3-003.
 64. Silén H, Salih EY, Mgbeahurike EE, Fyhrqvist P. Ethnopharmacology, antimicrobial potency, and phytochemistry of African *Combretum* and *Pteleopsis* species (Combretaceae): a review. *Antibiotics (Basel).* 2023;12(2):264. doi: 10.3390/antibiotics12020264, PMID 36830175.
 65. Long C. Swaziland's Flora: SiSwati names and uses. Swaziland National Trust commission, Mbambane; 2005. [cited Nov 3 2024] Available from: <http://www.sntc.org.sz/index.asp>.
 66. Cume D, Loon R, Bester D. Healing trees and plants of the Lowveld. Cape Town: Struik Nature; 2009.
 67. Chinsebu KC, Hjarunguru A, Mbangi A. Ethnomedicinal plants used by traditional healers in the management of HIV/AIDS opportunistic diseases in Rundu, Kavango East region, Namibia. *S Afr J Bot.* 2015;100:33-42. doi: 10.1016/j.sajb.2015.05.009.
 68. Cock IE, Van Vuuren SF. A comparison of the antimicrobial activity and toxicity of six *Combretum* and two *Terminalia* species from southern Africa. *Pharmacogn Mag.* 2015;11(41):208-18. doi: 10.4103/0973-1296.149740, PMID 25709234.
 69. Samie A, Madzie N. Effects of *Combretum hereroense* and *Canthium mundianum* water extracts on production and expression of interleukin-4. *Afr J Trad Compl Alt Med.* 2016;23;14(1):302-9. doi: 10.21010/ajtcam.v14i1.32.
 70. Isabel M, Jossias AD, Maria JS, Natasha R, Isabel IA. Woody species from the Mozambican miombo woodlands: a review on their ethnomedicinal uses and pharmacological potential. *J Med Plants Res.* 2018;12(2):15-31. doi: 10.5897/JMPR2017.6540.
 71. Tchamba JJ, Catarino S, Duarte MC, Silva JM, Romeiras MM, Catarino L. Ethnobotany in Angola: the contribution of late 19th century Christian missionaries to the knowledge of medicinal wild plants. *J Ethnopharmacol.* 2024;334:118511. doi: 10.1016/j.jep.2024.118511, PMID 38969150.
 72. Motlhanka DM, Nthoiwa GP. Ethnobotanical survey of medicinal plants of Tswapong North, in eastern Botswana: A case of plants from Mosweu and Seolwane villages. *Eur J Med Pl.* 2013;3(1):10-24. doi: 10.9734/EJMP/2013/1871.
 73. DEN Mabogo. The ethnobotany of the Vhavenda. Pretoria [masters dissertation]. University of Pretoria; 1990.
 74. Hutchings A, Scott AH, Lewis G, Cunningham AB. Zulu medicinal plants: an inventory. Pietermaritzburg: University of Natal Press; 1996.
 75. Chinemana F, Drummond RB, Mavi S, De Zoysa I. Indigenous plant remedies in Zimbabwe. *J Ethnopharmacol.* 1985;14(2-3):159-72. doi: 10.1016/0378-8741(85)90084-4, PMID 4094463.
 76. Samuelsson G, Farah MH, Claeson P, Hagos M, Thulin M, Hedberg O *et al.* Inventory of plants used in traditional medicine in Somalia. I. Plants of the families Acanthaceae-Chenopodiaceae. *J Ethnopharmacol.* 1991;35(1):25-63. doi: 10.1016/0378-8741(91)90132-w, PMID 1753795.
 77. Leffers A. Gemsbok bean and Kalahari truffle. Windhoek: Gamsberg MacMillan; 2003.
 78. Pakia M, Cooke JA, Van Staden J. The ethnobotany of the Midzichenda tribes of the coastal forest areas in Kenya: 2. Medicinal plant uses. *S Afr J Bot.* 2003;69(3):382-95. doi: 10.1016/S0254-6299(15)30321-5.
 79. Semanya SS, Maroyi A. Medicinal plants used for the treatment of tuberculosis by Bapedi traditional healers in three districts of the Limpopo province, South Africa. *Afr J Tradit Complement Altern Med.* 2013;10(2):316-23. doi: 10.4314/ajtcam.v10i2.17, PMID 24146456.
 80. Semanya SS, Maroyi A. Data on medicinal plants used to treat respiratory infections and related symptoms in South Africa. *Data Brief.* 2018;21:419-23. doi: 10.1016/j.dib.2018.10.012, PMID 30364644.
 81. Semanya SS, Maroyi A. Source, harvesting, conservation status, threats and management of indigenous plant used for respiratory infections and related symptoms in the Limpopo province, South Africa. *Biodivers J Biol Divers.* 2019;20(3):789-810. doi: 10.13057/biodiv/d200325.
 82. Semanya SS, Maroyi A. Source of plants, used by Bapedi traditional healers for respiratory infections and related symptoms in the Limpopo province, South Africa. *J Biol Sci.* 2019;19(2):101-21. doi: 10.3923/jbs.2019.101.121.
 83. Mutie FM, Gao LL, Kathambi V, Rono PC, Musili PM, Ngugi G *et al.* An ethnobotanical survey of a dryland botanical garden and its environs in Kenya: the Mutomo Hill plant sanctuary. *Evid-Based Compl Alt Med.* 2022;2020:Article ID 1543831. doi: 10.1155/2020/1543831.
 84. Mutie FM, Mbuni YM, Rono PC, Mkala EM, Nzei JM, Phumthum M, *et al.* Important medicinal and food taxa (orders and families) in Kenya, based on three quantitative approaches. *Plants (Basel).* 2023;12(5):1145. doi: 10.3390/plants12051145, PMID 36904005.
 85. Nyirenda J, Chipuwa M. An ethnobotanical study of herbs and medicinal plants used in western, Copperbelt, central and northern provinces of Zambia. *Phytomed Plus.* 2024;4(1):100514. doi: 10.1016/j.phyplu.2023.100514.
 86. Chinsebu KC. Ethnobotanical study of medicinal flora utilised by traditional healers in the management of sexually transmitted infections in Sesheke District, western province, Zambia. *Rev Bras Farmacog.* 2015;26:268-74. doi: 10.1016/j.bjp.2015.07.030.
 87. Chakale MV, Asong JA, Struwig M, Mwanza M, Aremu AO. Ethnoveterinary practices and ethnobotanical knowledge on plants used against cattle diseases among two communities in South Africa. *Plants (Basel).* 2022;11(13):1784. doi: 10.3390/plants11131784, PMID 35807736.
 88. Letcher RM, Nhamo LR. Chemical constituents of the Combretaceae. Part IV. Phenanthrene derivatives from the heartwood of *Combretum hereroense*. *J Chem Soc Perkin Trans 1.* 1973;1:1179-81. doi: 10.1039/P19730001179.
 89. Katerere DR, Gray AI, Nash RJ, Waigh RD. Phytochemical and antimicrobial investigations of stilbenoids and flavonoids isolated from three species of Combretaceae. *Fitoterapia.* 2012;83(5):932-40. doi: 10.1016/j.fitote.2012.04.011, PMID 22546149.
 90. Grimsey L, Van Vuuren SF, Wright MH, Cock IE. Selected South African *Combretum* spp. extracts inhibit methicillin-resistant *Staphylococcus aureus* and ESBL strains of *Escherichia coli* and *Klebsiella pneumoniae*. *S Afr J Bot.* 2024;165:49-58. doi: 10.1016/j.sajb.2023.12.018.
 91. McGaw LJ, Rabe T, Sparg SG, Jäger AK, Eloff JN, Van Staden J. An investigation on the biological activity of *Combretum* species. *J Ethnopharmacol.* 2001;75(1):45-50. doi: 10.1016/S0378-8741(00)00405-0, PMID 11282442.
 92. Eloff JN. The antibacterial activity of 27 southern African members of the Combretaceae. *S Afr J Sci.* 1999;95:148-52.
 93. Fyhrquist P, Mwasumbi L, Haeggström CA, Vuorela H, Hiltunen R, Vuorela P. Ethnobotanical and antimicrobial investigation on some species of *Terminalia* and *Combretum* (Combretaceae) growing in Tanzania. *J Ethnopharmacol.* 2002;79(2):169-77. doi: 10.1016/S0378-8741(01)00375-0, PMID 11801378.
 94. Anokwuru CP, Sandasi M, Chen W, Van Vuuren S, Elisha IL, Combrinck S *et al.* Investigating antimicrobial compounds in South African Combretaceae species using a biochemometric approach. *J Ethnopharmacol.* 2021;269:113681. doi: 10.1016/j.jep.2020.113681, PMID 33307052.
 95. Masoko P, Nxumalo KM. Validation of antimycobacterial plants used by traditional healers in three districts of the Limpopo province (South Africa). *Evid Based Complement Alternat Med.* 2013;2013:Article ID 586247. doi: 10.1155/2013/586247, PMID 23956776.
 96. Komape NP, Bagla VP, Kabongo-Kayoka P, Masoko P. Antimycobacteria potential and synergistic effects of combined crude extracts of selected medicinal plants used by Bapedi traditional healers to treat tuberculosis related symptoms in Limpopo province, South Africa. *BMC Complement Altern Med.* 2017;17(1):128. doi: 10.1186/s12906-016-1521-2, PMID 28235402.
 97. Masoko P, Picard J, Eloff JN. The antifungal activity of twenty-four southern African *Combretum* species (Combretaceae). *S Afr J Bot.* 2007;73(2):173-83. doi: 10.1016/j.sajb.2006.09.010.
 98. Eloff JN, Jäger AK, Van Staden J. The stability and relationship between anti-inflammatory activity and antibacterial properties of southern African *Combretum* species. *S Afr J Sci.* 2001;97:291-3.
 99. Masoko P, Eloff JN. Screening of twenty-four South African *Combretum* and six terminalia species (Combretaceae) for antioxidant activities. *Afr J Trad Compl Alt Med.* 2007;4(2):231-9. doi: 10.4314/ajtcam.v4i2.31213.