

**Review Article**
**HEPATOPROTECTIVE HERBS: AN UPDATED REVIEW**
**KUNAL WALIA\*, AJEET PAL SINGH, AMAR PAL SINGH**

Department of Pharmacology, St. Soldier Institute of Pharmacy, Lidhran Campus, Behind NIT (R. E. C.), Jalandhar–Amritsar by pass, NH-1, Jalandhar-144011, Punjab, India

 \*Corresponding author: Kunal Walia; \*Email: [kunalwalia2000.k@gmail.com](mailto:kunalwalia2000.k@gmail.com)

Received: 10 Jun 2025, Revised and Accepted: 02 Aug 2025

**ABSTRACT**

Liver diseases are becoming a serious global health problem and may be caused by many toxic substances, including chemotherapeutic drugs, thioacetamide, carbon tetrachloride, certain antibiotics, excessive alcohol consumption, and microbes. Therefore, having a healthy liver is vitally important for good health and well-being. The liver is an important organ, contributing to the metabolism of the body and that of xenobiotics. There are many toxic substances which could cause liver damage (certain antibiotics, chemotherapeutic drugs, carbon tetrachloride, thioacetamide, and microbes, are mostly responsible for liver cell damage). The synthetic drugs that are available at this point to deal with liver dysfunction causes additional harm to the liver. Hence, the use of herbal medicines has grown and become popular. The important medicinal herbs that can be used to treat liver diseases with the least impact on the kidneys have been described. Because of the hepatoprotective character, antioxidant-related characteristics and least harm to kidneys, features of the newly described medicinal plants can be used to develop new medicines for the prevention and treatment of liver diseases.

**Keywords:** Herbal drugs, Hepatotoxicity, Liver diseases, Medicinal plant, Factors

 © 2025 The Authors. Published by Innovare Academic Sciences Pvt Ltd. This is an open access article under the CC BY license (<https://creativecommons.org/licenses/by/4.0/>) DOI: <https://dx.doi.org/10.22159/ijcpr.2025v17i5.7045> Journal homepage: <https://innovareacademics.in/journals/index.php/ijcpr>
**INTRODUCTION**

With an average adult weight of 1.4 kg, the liver is the heaviest gland in the human body (after the skin) and the second largest organ. It occupies the majority of the right hypochondriac and a portion of the epigastric regions of the abdominopelvic cavity and is located inferior to the diaphragm [1]. The liver plays a major role in the metabolism and excretion of drugs. Detoxifying pharmaceuticals and xenobiotics in the liver via drug metabolizing enzymes (DMEs) is one of the significant processes involved in the restoration of homeostasis [2]. The liver accounts for 2 % to 3 % of normal body weight. It normally has four lobes, each defined in both morphological and functional anatomy. It is located underneath the right hemidiaphragm (right upper quadrant of the abdominal cavity) and has ligamentous attachments to the area. It is protected by the ribcage and is held in place by peritoneal reflections. Although the liver's ligamentous attachments are not true ligaments, they are avascular and continuous with the Glisson capsule or the liver's visceral peritoneum [3]. Hepatotoxicity refers to damage to the liver caused by chemicals. Drug-induced damage to the liver can cause acute and chronic liver disease. The liver is susceptible to the toxicity wrought by many other chemicals, and the liver's role to metabolize and excrete toxins is of paramount importance. Several drugs, including acetaminophen, can lead to harm to an organ by overtaking otherwise modest therapeutic dosages. The liver can also be damaged by other chemical agents, e. g., in factories or laboratories, naturally occurring compounds (e. g., microcystins), and herbal medicines. Substances that are harmful to the liver are also termed hepatotoxins [4]. The most common reason for drugs to be withdrawn from the market is liver damage; drug-induced liver damage has been linked to 900+ drugs, and hepatotoxicity and drug-

induced liver injury are also responsible for the failure of many compounds that are under investigation so that it is essential that new drug screening assays could use new stem cell-derived hepatocyte-like cells, which will determine hepatotoxicity much earlier in the drug developmental process. Subclinical liver damage occurs with the administration of chemicals and will typically only manifest as abnormal liver enzymes. Drug-induced liver damage is responsible for 50% of all cases of acute liver failures and 5% of all hospital admissions [5]. Most of these substances are absorbed through the gastrointestinal tract, but a smaller number though parenterally or absorbed through the skin or lungs immediately (inhalants). The majority of drugs and xenobiotics are lipophilic because they pass readily through the intestine and hepatocyte cell membranes. Metabolism in the hepatocyte alters drug structure in such a way that the drug is more hydrophilic, producing water soluble metabolites which are cleared in urine or bile. Drug metabolic pathways are divided into phase I pathways (oxidation, reduction, hydrolysis) mediated by various cytochrome P450 isozymes and phase II conjugation pathways (glucuronidation, acetylation, sulphation, methylation) mediated by various transferases. Highly polar drugs do not require metabolism. Some drugs may degrade spontaneously [6].

**Genetic and nongenetic risk factors**

Risk factors for susceptibility to drug-induced hepatotoxicity

- Toxic potential of drug: such as Reactive Metabolites, Acyl glucuronide, Mitochondrial effects
- Genetic factors such as drug metabolism, detoxification, transport
- Environmental factors such as ethanol and Age/Sex

**Table 1: Therapeutic agents causing hepatotoxicity [7, 8]**

Antimicrobial	Analgesics and anti-tuberculosis drug	Immunomodulator	Antiepileptics
Amoxicillin	NSAIDs	Interferon-beta	Phenytoin
Isoniazid	Rifampicin, Rifabutin	Interferon-alpha	Lamotrigine
Sulfamethoxazole	Pyrazinamide	Anti-TNF agents Azathioprine	Valproic Acid
Trimethoprim	Prothionamide	Cyclophosphamide	Carbamazepine

Table 2: Medicinal plants with hepatoprotective potentials

Name of the plant	Extract used	Hepatotoxicity-inducing agents	Biochemical and histopathological parameter studied	Hepatotoxicity-inducing agents
<i>Phyllanthus Muellarianus</i> (leaves) [9]	Aqueous	ALP, ALT, AST, ALB, TB, CAT	Acetaminophen	400 mg/kg
<i>Picrorhizakurroa</i> (Roots) [10]	Ethanol	SGOT, SGPT, ALP	CCI4	2.60 ml/kg
<i>Bauhinia Variegata</i> (Stem barks) [11]	Alcohol	AST, ALP, GGT, ALT	CCI4	100 and 200 mg/kg
<i>Galium aparine</i> (whole) [12]	Alcohol	ALT, AST, and ALP	CCI4	2 ml/kg
<i>Ficus cordata</i> (roots) [13]	Methanol/ethyl	LDH	CCI4	400 mg/kg
<i>Canna indica</i> (Aerial parts) [14]	Methanol	SGPT, SGOT, TB, CAT, GSH, LPO	CCI4	100 and 200 mg/kg
<i>Curcuma longa</i> (Rhizome)[15]	Ethanol	ALT, ALP, and AST	PCM	600 mg/kg
<i>Dodonaeaviscosa</i> (leaves) [16]	Methanol	AST, LDLC, ALT	Alloxan	500 mg/kg
<i>Ecliptaprostrate</i> (Fresh leaves)[17]	Methanol	ALT, AST, and serum bilirubin	CCI4	10 80 mg/kg
<i>Boerhaviadiffusa</i> (Roots)[18]	Ethanol	SGPT, SAP, TGs, and total lipid levels	Country-made liquor	200 and 400 mg/kg
<i>Tylophora</i> (leaves) [19]	Methanol	SGPT, ALP, SGOT	CCI4	200 and 300 mg/kg
<i>Tylophora</i> (Leaves) [20]	Methanol	SGPT, ALP, SGOT	CCI4	200 and 300 mg/kg
<i>Tridax Procumbens</i> (Aerial parts) [21]	Ethanol	AST, LDH, ALT, ALP, GGT, TB	d-GalN/IPS	300 mg/kg
<i>Opuntia ficus-indica</i> (Leaves) [22]	Aqueous	AST, ALT, Creatinine, Urea, Uric acid	CCI4	2, ml/kg
<i>Apium graveolens</i> (Seeds) [23]	Methanol	SGOT, SGPT, SALP	CCI4	250 mg/Kg
<i>Opuntia ficus-indica</i> (Stem) [24]	Aqueous	ALAT, ASAT, ALP, LDH, CHL	CPF	1500 mg/kg
<i>Agrimoniaeupatoria</i> (Aerial part) [25]	Aqueous	AST and ALT	Ethanol	100 and 300 mg/kg
<i>Vitis vinifera</i> (Leaves) [26]	Alcohol	AST and ALT	CCI4	125 mg/kg
<i>Rheum palmatum</i> (Aerial part) [27]	N/A	N/A	CCI4/ethanol	25 and 100 mg/kg
<i>Ziziphus oenoplia</i> (Roots) [28]	Alcohol	SGOT, SGPT, SALP, SB, SOD, CAT, GST	INH and RIF	150 and 300 mg/kg
<i>Corylus avellana</i> (Leaves) [29]	Aqueous	GPT and GOT	CCI4 and acetaminophen	NA
<i>Cinnamomum Cassia</i> (Bark) [30]	Ethanol	TP, albumin, TB, direct bilirubin,	Dimethyl nitrosamine	40 mg/kg
<i>Pistacia lentiscus</i> (Gums) [31]	NA	AST, ALT and MDA, GSH, GPx, GST, GR, SOD	CCI4	NA
<i>Punica granatum</i> (Edible and Portion) [32]	Acetone	AST, ALT, and LDH	INH and RIF	400 mg/kg
<i>Rosa damascene</i> Mill (Flower) [33]	Aqueous	AST, ALT, ALP, LDH, ALBTB, urea and creatinine	Acetaminophen	250, 500 and 1000 mg/kg
<i>Cucurbita maxima</i> (Aerial parts) [34]	Methanol	SGPT, SGOT, ALP, TP, and TB	CCI4	250 and 500 mg/kg
<i>Cynara</i> (Root) [35]	Hydroalcohol	ALT, ALP, AST,	CCI4	900 mg/kg
<i>Taraxacum Officinale</i> (Roots) [36]	Hydroalcoholic acid	TBARS, GST, GSH, SOD, CAT, GR, and	Ethanol	250 mg/kg
<i>Tragopogon Porrifolius</i> (Edible root and shoot) [37]	Methanol	CAT, SOD and GSTAST, ALT	CCI4	250 mg/kg
<i>Baliospermum Montanum</i> (Root) [38]	Methanol	GOT, GPT, ALP, TB, TC, TB	TAA	2000 mg/kg
<i>Tephrosia Purpurea</i> (Aerial parts) [39]	Ethanol	AST, GSH, ALT, ALP, TB, GGT	TAA	500 mg/kg
<i>Alchornea Cordifolia</i> (leaves) [40]	Methanol	SGOT/AST, SGPT/ALT, ALP	CCI4	300 mg/kg
<i>Glycosmis pentaphylla</i> Corr. (Leaves, bark) [41]	Methanol	ALT/SGPT, AST/SGOT, CHL	CCI4	500 mg/kg
<i>Wedelia chinensis</i> L. (Leaves) [42]	Ethanol	AST, ALT, ALP, Protein	CCI4	200 mg/kg
<i>Cassia fistula</i> (Seeds) [43]	Methanol	SGOT, SGPT, ALP, and bilirubin	PCM	200 and 400 mg/kg
<i>Tylophora</i> (Leaves) [19]	Methanol	SGPT, ALP, SGOT	CCI4	200 and

## CONCLUSION

New hepatoprotective drugs need to be discovered because current medications that treat liver diseases, particularly a viral hepatitis or any degree of chronic liver disease, do not serve the unmet needs of the patients and may have detrimental renal effects. Because liver function is of utmost importance to the human body, liver disease and liver injury is now ranked as one of the most impotent health issues in the world. The causes of liver injury primarily include excessive intake of alcoholic beverages, lack of dietary discipline, herbal supplement use, viral, bacterial and parasitic infections, autoimmune diseases, neoplastic processes, metabolic disorders, and substance abuse. For example 50% of people living in developing countries that have liver disease or any form of liver disease choose to use herbal medicine for treatment and therefore have tapped worldwide interest. The majority of herbal extracts that are available today have demonstrated great effect with mild effects on regulating the signs and symptoms associated with liver disease or liver injury. In the treatment of liver disease or the symptoms associated with liver disease, the herbs may have provided a new avenue to explore in the constrained options for medicinal measures.

## ACKNOWLEDGMENT

It's our privilege to express the profound sense of gratitude and cordial thanks to our respected chairman Mr. Anil Chopra and Vice Chairperson, Ms. Sangeeta Chopra, St. Soldier Educational Society, Jalandhar for providing the necessary facilities to complete this review work.

## FUNDING

Nil

## AUTHORS CONTRIBUTIONS

All authors have contributed equally

## CONFLICT OF INTERESTS

Declared none

## REFERENCES

1. Tortora GJ, Derrickson B. Anatomy and physiology. John Wiley & Sons India Pvt Limited; 2014.

2. Singh D, Cho WC, Upadhyay G. Drug-induced liver toxicity and prevention by herbal antioxidants: an overview. *Front Physiol.* 2015;6:363. doi: [10.3389/fphys.2015.00363](#), PMID [26858648](#).
3. Abdel Misih SR, Bloomston M. Liver anatomy. *Surg Clin North Am.* 2010 Aug;90(4):643-53. doi: [10.1016/j.suc.2010.04.017](#), PMID [20637938](#), PMCID [PMC4038911](#).
4. Pandit A, Sachdeva T, Bafna P. Drug induced hepatotoxicity: a review. *J Appl Pharm Sci.* 2012 May 30;233-43. doi: [10.7324/JAPS.2012.2541](#).
5. Highland HN, Rishika AS, Almira SS, Kanthi PB. Ficoll-400 density gradient method as an effective sperm preparation technique for assisted reproductive techniques. *J Hum Reprod Sci.* 2016 Jul 1;9(3):194-9. doi: [10.4103/0974-1208.192070](#), PMID [27803588](#).
6. Rastogi S, Arora P, Kapoor S, Wazir SS, Vashishth S, Sharma V. Prevalence of oral soft tissue lesions and medical assessment of geriatric outpatients in North India. *J Indian Acad Oral Med Radiol.* 2015 Jul 1;27(3):382-6. doi: [10.4103/0972-1363.170461](#).
7. Chalasani N, Younossi Z, Lavine JE, Charlton M, Cusi K, Rinella M. The diagnosis and management of nonalcoholic fatty liver disease: practice guidance from the American association for the study of liver diseases. *Hepatology.* 2018 Jan;67(1):328-57. doi: [10.1002/hep.29367](#), PMID [28714183](#).
8. Ozougwu JC. Anti-diabetic effects of Allium cepa (onions) aqueous extracts on alloxan induced diabetic Rattus norvegicus. *J Med Plants Res.* 2011 Apr 4;5(7):1134-9. doi: [10.5897/JMPR.9000094](#).
9. Ajiboye TO, Ahmad FM, Daisi AO, Yahaya AA, Ibitoye OB, Muritala HF. Hepatoprotective potential of Phyllanthus muellarianus leaf extract: studies on hepatic oxidative stress and inflammatory biomarkers. *Pharm Biol.* 2017 Jan 1;55(1):1662-70. doi: [10.1080/13880209.2017.1317819](#), PMID [28447517](#).
10. Arsul VA, Wagh SR, Mayee RV. Hepatoprotective activity of liver gen a polyherbal formulation against carbon tetrachloride induced hepatotoxicity in rats. *Int J Pharm Pharm Sci.* 2011;3(3):228-31.
11. Ajay KG, Neelam M. Hepatoprotective activity of aqueous and ethanolic extract of Boerhaavia erecta. In carbon tetrachloride intoxicated albino rats. *Am J Pharmacol Toxicol.* 2006;1:17-20.
12. Khan MU, Rohilla A, Bhatt D, Afrin S, Rohilla S, Ansari SH. Diverse belongings of Calendula officinalis: an overview. *Int J Pharm Sci Drug Res.* 2011;3(3):173-7. doi: [10.25004/IJPSDR.2011.030302](#).
13. Donfack HJ, Kengap RT, Ngameni B, Chuisseu PD, Tchana AN, Buonocore D. Ficus cordata Thunb. (Moraceae) is a potential source of some hepatoprotective and antioxidant compounds. *Pharmacologia.* 2011;2(5):137-45. doi: [10.5567/pharmacologia.2011.137.145](#).
14. Arulkumaran KS, Rajasekaran A, Ramasamy A, Jegadeesan M, Kavamani S, Somasundaram A. Amaranthus spinosus. seeds protect liver against toxic effects of ethanol and carbon tetrachloride in rats. *Int J Pharm Tech Res.* 2009;1:273-46.
15. Hsieh CC, Fang HL, Lina WC. Inhibitory effect of Solanum nigrum on thioacetamide induced liver fibrosis in mice. *J Ethnopharmacol.* 2008;119(1):117-21. doi: [10.1016/j.jep.2008.06.002](#), PMID [18606216](#).
16. Ahmed A, Shah WA, Akbar S, Kumar D, Kumar V, Younis M. *In vitro* anti-inflammatory activity of Salix caprae Linn. (goat willow) by HRBC membrane stabilization method. *J Pharm Res.* 2011;4:1067-8. doi: [10.13140/RG.2.2.17974.80965](#).
17. Lin HM, Tseng HC, Wang CJ, Lin JJ, Lo CW, Chou FP. Hepatoprotective effects of Solanum nigrum Linn extract against CCl<sub>4</sub>-induced oxidative damage in rats. *Chem Biol Interact.* 2008;171(3):283-93. doi: [10.1016/j.cbi.2007.08.008](#), PMID [18045581](#).
18. Mishra G, Khosa RT, Singh P, Jha KK. Hepatoprotective potential of ethanolic extract of Caesalpinia crista leaves against paracetamol induced hepatotoxicity in rats. *J Coast Life Med.* 2014;2:575-9.
19. Mujeeb M, Aeri V, Bagri P, Khan SA. Hepatoprotective activity of methanolic extract of Tylophora indica (Burm. F.) Merrill. leaves. *Int J Green Pharm.* 2010;3(2):125-7. doi: [10.4103/0973-8258.54901](#).
20. Singh MK, Sahu P, Nagori K, Dewangan D, Kumar T, Alexander A. Organoleptic properties *in vitro* and *in vivo* pharmacological activities of calendula officinalis linn: an over review. *J Chem Pharm Res.* 2011;3(4):655-63.
21. Meena B, Ezhilan RA, Rajesh R, Hussain KS, Ganesan B, Anandan R. Antihepatotoxic potential of Tridax procumbens on antioxidant defense status in D-galactosamine induced hepatitis in rats. *Afr J Biochem Res.* 2008;2(2):51-5.
22. Djerrou Z, Maameri Z, Halmi S, Djaalab H, Riachi F, Benmaiza L. Hepatoprotective effect of Opuntia ficus-indica aqueous extract against carbon tetrachloride induced toxicity in rats. *Online J Biol Sci.* 2015 Apr 1;15(2):36-41. doi: [10.3844/ojbsci.2015.36.41](#).
23. Ahmed B, Alam T, Varshney M, Khan SA. Hepatoprotective activity of two plants belonging to the Apiaceae and the Euphorbiaceae family. *J Ethnopharmacol.* 2002;79(3):313-6. doi: [10.1016/S0378-8741\(01\)00392-0](#), PMID [11849834](#).
24. Ncibi S, Ben Othman M, Akacha A, Krifi MN, Zourgui L. Opuntia ficus indica extract protects against chlorpyrifos induced damage on mice liver. *Food Chem Toxicol.* 2008;46(2):797-802. doi: [10.1016/j.fct.2007.08.047](#), PMID [17980473](#).
25. Yoon SJ, Koh EJ, Kim CS, Zee OP, Kwak JH, Jeong WJ. Agrimonia eupatoria protects against chronic ethanol induced liver injury in rats. *Food Chem Toxicol.* 2012 Jul 1;50(7):2335-41. doi: [10.1016/j.fct.2012.04.005](#), PMID [22525864](#).
26. Orhan DD, Orhan N, Ergun E, Ergun F. Hepatoprotective effect of Vitis vinifera L. leaves on carbon tetrachloride induced acute liver damage in rats. *J Ethnopharmacol.* 2007;112(1):145-51. doi: [10.1016/j.jep.2007.02.013](#), PMID [17391882](#).
27. Guo MZ, Li XS, Xu HR, Mei ZC, Shen W, Ye XF. Rhein inhibits liver fibrosis induced by carbon tetrachloride in rats. *Acta Pharmacol Sin.* 2002 Aug 1;23(8):739-44. PMID [12147197](#).
28. Rao CHV, Rawat AK, Singh AP, Singh A, Verma N. Hepatoprotective potential of ethanolic extract of Ziziphus oenoplia (L.) mill roots against antitubercular drugs induced hepatotoxicity in experimental models. *Asian Pac J Trop Med.* 2012;5(4):283-8. doi: [10.1016/S1995-7645\(12\)60040-6](#), PMID [22449519](#).
29. Rusu MA, Bucur N, Puica C, Tamas M. Effects of Corylus avellana in acetaminophen and CCl<sub>4</sub> induced toxicosis. *Phytother Res.* 1999;13(2):120-3. doi: [10.1002/\(SICI\)1099-1573\(199903\)13:2<120::AID-PTR403>3.0.CO;2-I](#), PMID [10190184](#).
30. Lim CS, Kim EY, Lee HS, Soh Y, Sohn Y, Kim SY. Protective effects of Cinnamomum cassia Blume in the fibrogenesis of activated HSC-T6 cells and dimethylnitrosamine induced acute liver injury in SD rats. *Biosci Biotechnol Biochem.* 2010 Mar 23;74(3):477-83. doi: [10.1271/bbb.90435](#), PMID [20208363](#).
31. Mavridis SK, Gortzi O, Lalas S, Paraschos S, Skaltsounis AL, Pappas IS. Hepatoprotective effect of Pistacia lentiscus var. Chia total extract against carbon tetrachloride induced liver damage in rats. *Planta Med.* 2008;74(9). doi: [10.1055/s-0028-1084336](#).
32. Yogeeta S, Ragavender HR, Devaki T. Antihepatotoxic effect of Punica granatum. Acetone extract against isoniazid and rifampicin induced hepatotoxicity. *Pharm Biol.* 2007;45(8):631-7. doi: [10.1080/13880200701538963](#).
33. Saxena M, Shakya AK, Sharma N, Shrivastava S, Shukla S. Therapeutic efficacy of Rosa damascena mill. on acetaminophen induced oxidative stress in albino rats. *J Environ Pathol Toxicol Oncol.* 2012;31(3):193-201. doi: [10.1615/jenvironpatholtoxiconcol.v31.i3.10](#), PMID [23339694](#).
34. Saha P, Mazumder UK, Haldar PK, Bala A, Kar B, Naskar S. Evaluation of hepatoprotective activity of Cucurbita maxima aerial parts. *J Herb Med Toxicol.* 2011;5(1):17-22.
35. Fallah HH, Zareei MA, Ziai SA, Mehrzama M, Alavian SM, Mehdizadeh M. The effects of Cynara scolymus L. leaf and Cichorium intybus L. root extracts on carbon tetrachloride induced liver toxicity in rats. *Journal of Medicinal Plants.* 2011 Mar;10(37):33-9.
36. Das SK, Mukherjee S. Biochemical and immunological basis of silymarin effect a milk thistle (Silybum marianum) against ethanol induced oxidative damage. *Toxicol Mech Methods.* 2012;22(5):409-13. doi: [10.3109/15376516.2012.673090](#), PMID [22409310](#).
37. Mroueh M, Daher C, El Sibai M, Tenkerian C. Antioxidant and hepatoprotective activity of Tragopogon porrifolius methanolic extract. *Planta Med.* 2011;77(12). doi: [10.1055/s-0031-1282460](#).

38. Kumar SV, Mishra SH. Hepatoprotective effect of *Baliospermum montanum* (Willd.) Muell. arg against thioacetamide induced toxicity. Int J Compreh Pharm. 2012;3(9):1-4.
39. Khatri A, Garg A, Agrawal SS. Evaluation of hepatoprotective activity of aerial parts of *Tephrosia purpurea* L. and stem bark of *Tecomella undulata*. J Ethnopharmacol. 2009;122(1):1-5. doi: [10.1016/j.jep.2008.10.043](https://doi.org/10.1016/j.jep.2008.10.043), PMID [19059328](https://pubmed.ncbi.nlm.nih.gov/19059328/).
40. Osadebe PO, Okoye FB, Uzor PF, Nnamani NR, Adiele IE, Obiano NC. Phytochemical analysis hepatoprotective and antioxidant activity of *alchornea cordifolia* methanol leaf extract on carbon tetrachloride induced hepatic damage in rats. Asian Pac J Trop Med. 2012 Apr 1;5(4):289-93. doi: [10.1016/S1995-7645\(12\)60041-8](https://doi.org/10.1016/S1995-7645(12)60041-8), PMID [22449520](https://pubmed.ncbi.nlm.nih.gov/22449520/).
41. Ahsan MR, Islam KM, Bulbul IJ. Hepatoprotective activity of methanol extract of some medicinal plants against carbon tetrachloride induced hepatotoxicity in rats. Eur J Sci Res. 2009;37(2):302-10.
42. Murugaian P, Ramamurthy V, Karmegam N. Hepatoprotective activity of *Wedelia calendulacea* L. against acute hepatotoxicity in rats. Res J Agric Biol Sci. 2008;4:685-7.
43. Chaudhari NB, Chittam KP, Patil VR. Hepatoprotective activity of *Cassia fistula* seeds against paracetamol induced hepatic injury in rats. Arch PharmSci Res. 2009;1:218-21.